

Tetrahedron Letters 43 (2002) 9489-9492

TETRAHEDRON LETTERS

Spirocyclization strategy toward indole phytoalexins. The first synthesis of (±)-1-methoxyspirobrassinin, (±)-1-methoxyspirobrassinol, and (±)-1-methoxyspirobrassinol methyl ether

Peter Kutschy,^{a,*} Mojmír Suchý,^a Kenji Monde,^b Nobuyuki Harada,^c Renata Marušková,^a Zuzana Čurillová,^a Milan Dzurilla,^a Mariana Miklošová,^a Roman Mezencev^a and Ján Mojžiš^d

^aInstitute of Chemical Sciences, Faculty of Science, P. J. Šafárik University, Moyzesova 11, 041 67 Košice, Slovak Republic ^bDivision of Biological Sciences, Graduate School of Science, Hokkaido University, Kita 10, Nishi 8, Sapporo 060-0810, Japan ^cInstitute of Multidisciplinary Research for Advanced Materials, 2-1-1, Katahira, Aoba-ku, Sendai 980-8577, Japan ^dInstitute of Pharmacology, Faculty of Medicine, P. J. Šafárik University, SNP 1, 040 66 Košice, Slovak Republic

Received 16 September 2002; revised 22 October 2002; accepted 1 November 2002

Abstract—The first syntheses of cruciferous indole phytoalexins (\pm) -1-methoxyspirobrassinin, (\pm) -1-methoxyspirobrassinol methyl ether as well as a new syntheses of phytoalexins (\pm) -spirobrassinin and cyclobrassinin were achieved by dioxane dibromide (DDB)-mediated spirocyclization of brassinin and its 1-substituted derivatives. © 2002 Elsevier Science Ltd. All rights reserved.

Phytoalexins are defined as antimicrobial low molecular weight secondary metabolites, produced by plants after their exposure to physical, biological or chemical stress.¹ About 30 indole phytoalexins were isolated from the economically and dietary important plants of the family *Cruciferae*, cultivated worldwide.² Selected examples (except for the synthetic analogs **7**, **8** and **11**) are shown in Fig. 1. With respect to their interesting biological properties, including antitumor activity³⁻⁶ it is important to develop synthetic methods toward cruciferous phytoalexins, since the isolation from plants does not afford sufficient quantities for biological screening.

In the present paper we report the preparation and antiproliferative activity of hitherto synthetically unavailable spiroindoline phytoalexins 3–5 and a new approach to the compounds 1 and 10. (S)-(–)-Spirobrassinin 1 was isolated in 1987 from Japanese radish,⁷ however, its synthesis by cyclization of dioxibrassinin 2^8 with SOCl₂ or MsCl and absolute stereochemistry were fully described only recently.⁹ (–)-1-Methoxyspirobrassinin (3) isolated in 1994 from kohlrabi¹⁰ and (-)-1methoxyspirobrassinol methyl ether 4 as well as optically inactive methoxyspirobrassinol 5, isolated in 1995 as a minor phytoalexin from Japanese radish¹¹ have not been synthesized to date and their biological properties are unknown. Compound 5 exists in solution as a mixture of diastereomers 5a and 5b in a 4:1 ratio, owing to its unstable hemiaminal structure.¹¹ The absolute stereochemistries of 3-5 remains unknown, formulae 4 and 5 show the diastereomeric structures only, as determined by NOE experiments.¹¹ Phytoalexins 3-5, 9 and 12 belong to a group of hydroxyindole natural products.¹² Our approach to the target compounds is based on the finding, that spirobrassinin 1 and cyclobrassinin 10 are synthesized in plants from brassinin 6^{13} Whereas 10 was prepared by cyclization of 6 with pyridinium tribromide¹⁴ or NBS⁴ in 34–35% yield and sinalbin B 12 was obtained by treatment of 1-methoxybrassinin 9 with NBS (41%),¹⁵ no transformation of brassinin 6 or its 1-substituted derivatives to spiroindoline phytoalexins has been reported to date.

Retrosynthetic analysis of 1 and 3–5 revealed a central role for a spiroindoleninium intermediate 13 (Scheme 1), which can undergo a rearrangement to the cyclobrassinin structure, or nucleophilic addition of water and methanol to a spiroindoline structure.

Keywords: phytoalexins; indoles; dioxane dibromide; spirocyclization; antitumor activity.

^{*} Corresponding author. Tel.: +421-55-62-22610, ext. 192; fax: +421-55-62-22124; e-mail: kutschy@kosice.upjs.sk

^{0040-4039/02/\$ -} see front matter @ 2002 Elsevier Science Ltd. All rights reserved. PII: S0040-4039(02)02452-8



Figure 1.





The preferred pathway should be dependent on the reactivity of iminium intermediate 13, towards nucleophiles. A low reactivity is expected to produce cyclobrassinin, and on the other hand its high reactivity should result in nucleophilic trapping resulting in the formation of spiroindolines.

To verify this prediction, brassinin 6^{14} and 1-methylbrassinin 7¹⁶ with low reactivity of the iminium intermediate versus 1-Boc-brassinin $\mathbf{8}^{16}$ with expected high reactivity of the corresponding acyliminium ion were selected as a model compounds. After several unsuccessful experiments with NBS, which seems to react by a different mechanism, involving bromination at the indole 3-position, we have found that dioxane dibromide (DDB) in dioxane in the presence of water (0.5%)is a suitable reagent for generation and rearrangement or trapping of iminium intermediate 13. The reaction probably starts at the thiocarbamoyl functionality with the formation of a sulfenyl bromide in which the electrophilic sulfur attacks the 3-position of indole to provide an iminium intermediate (Scheme 2). As anticipated, the iminium ions with R = H and CH_3 afforded cyclobrassinin 10 (45%) and 1-methylcyclobrassinin 11 (40%) identical with previously described compounds,^{14,16} whereas the acyliminium ion generated from the 1-Boc derivative 8 afforded a diastereomeric mixture of 1-Boc spirobrassinols 14a and 14b in a ratio 4:1 in 47% yield. Individual diastereomers were separated by flash chromatography and their structure determined on the basis of NOE-difference NMR experiments.¹⁷ However, under anhydrous conditions, 1-Boc-brassinin 8 reacted with the formation of cyclobrassinin 10 (45%), probably because the HBr liberated during the reaction deprotected the 9-Boc-cyclobrassinin, formed by the rearrangement of Boc-substituted intermediate 13. Compound 8 was consumed within 5 min and if, within the next 5 min, 0.5% of water was added, the spirobrassinols 14a and 14b were formed. However, when stirring was continued under anhydrous conditions, slow rearrangement and deprotection took place. We have found that the clean reaction with water as a nucleophile proceeds with up to 5% of water contained in the reaction medium. Therefore all further experiments were performed in dioxane/ water or dioxane/methanol 95:5 as the solvent.

A mixture of 14a and 14b was oxidized with CrO_3 to 1-Boc-spirobrassinin (±)-15, which after deprotection with TFA afforded spirobrassinin (±)-1. Treatment of a mixture of 14a and 14b with TFA proceeded with deprotection and water elimination toward an iminium cation with a trifluoroacetate counterion, which smoothly rearranged to cyclobrassinin 10 (Scheme 2).

Next, with the experimental conditions in hand for the preparation of 1-Boc-spirobrassinol 14, we attempted the cyclization of 1-methoxybrassinin 9.^{15,18,19} It appeared that the methoxyiminium intermediate formed is a highly reactive species, probably because of the activating effect of the electronegative oxygen atom. Consequently, the water present in the reaction mixture smoothly added to the carbon atom of the iminium intermediate to produce methoxyspirobrassinol 5 in 90% yield (Scheme 3).

Oxidation of 5 with CrO_3 afforded racemic 1methoxyspirobrassinin (±)-3 in 40% yield. If methanol was added as a nucleophile, instead of water, the methoxyiminium intermediate reacted with the formation of a mixture of the natural 4a and unnatural 4b



Scheme 3.

Scheme 2.

diastereomers of methoxyspirobrassinol methyl ether in a ratio of 1:2 which separated by flash chromatography (ether/hexane, 1:2). Spectroscopic data of the synthesized phytoalexins **1**, **3–5** and **10** are identical with those of the natural products.^{7,10,11,14}

Experimental procedure for 1-methoxyspirobrassinin: (\pm)-*Methoxyspirobrassinol* **5**: To a stirred solution of 1-methoxybrassinin **9** (63 mg, 0.24 mmol) in a mixture of dioxane/water (95:5, 5 mL) was added a freshly prepared solution of DDB (2 mL, 0.26 mmol; the stock solution was obtained by dissolving of 0.04 mL of bromine in 6 mL of dioxane). The reaction mixture was stirred for 10 min at room temperature, then triethylamine (0.26 mmol) was added, the mixture poured into water (50 mL), the product extracted with diethyl ether (2×10 mL), the extract washed with brine (20 mL), then dried over Na₂SO₄ and the residue obtained after evap-

oration of the solvent subjected to flash chromatography on 5 g of silica gel (cyclohexane/ethyl acetate, 5:1), affording 60 mg (90%) of 5 as a semi-solid colourless material with all the spectral data fully identical with the described natural product.¹¹ (\pm) -1-Methoxyspirobrassinin 3: To a stirred solution of 5 (91 mg, 0.32 mmol) in 98% acetic acid (9 mL) was added CrO₃ (160 mg, 1.6 mmol) and the mixture stirred for 1 h at room temperature. After pouring into water (40 mL), the product was extracted with diethyl ether $(2 \times 30 \text{ mL})$, the extract washed with 10% NaOH (2×15 mL) and brine (15 mL), dried over Na_2SO_4 and the residue obtained after evaporation of the solvent subjected to flash chromatography on 30 g of silica gel (diethyl ether/light petroleum, 1:2) to afford 36 mg (40%) of 3 as colourless needles, mp 129–131°C (ethyl acetate/hexane). The spectral data was fully identical with those of the natural product.¹⁰

The antiproliferative activity of the synthesized compounds was examined by MTT (thiazolyl blue) test,²⁰ using the selected human cancer cell lines (MDA-MB-231, MCF-7, U-87 MG and CACO-2). The highest activity was found with the natural diastereomer of methoxyspirobrassinol methyl ether **4a** which at concentration of 0.2 µmol L⁻¹ inhibited the growth of CACO-2 (colon adenocarcinoma) cell line to 38% of solvent control after 72 h incubation. This finding is consistent with the recommendation of consuming brassica vegetables as a protection against cancer.²¹

Acknowledgements

We thank the Slovak Ministry of Education (MVTS project Jap/Slov/UPJŠ) for financial support of this work.

References

- Purkayastha, R. P. In *Handbook of Phytoalexin* Metabolism and Action; Daniel, M.; Purkayastha, R. P., Eds. Progress in phytoalexin research during the past 50 years; Marcel Dekker: New York, 1995; pp. 1–39.
- For a recent review on cruciferous indole phytoalexins see: Pedras, M. S. C.; Okanga, F. I.; Zaharia, I. L.; Khan, A. Q. *Phytochemistry* 2000, 53, 161–176.
- Mehta, R. G.; Liu, J.; Constantinou, A.; Hawthorne, M.; Pezzuto, J. M.; Moon, R. C.; Moriarty, R. M. *Anticancer Res.* 1994, 14, 1209–1214.
- Mehta, R. G.; Liu, J.; Constantinou, A.; Thomas, C. F.; Hawthorne, M.; You, M.; Gerhäuser, C.; Pezzuto, J. M.; Moon, R. C.; Moriarty, R. M. *Carcinogenesis* 1995, 16, 399–404.
- Sabol, M.; Kutschy, P.; Siegfried, L.; Miroššay, A.; Suchý, M.; Hrbková, H.; Dzurilla, M.; Marušková, R.; Starková, J.; Paulíková, E. *Biologia (Bratislava)* 2000, 55, 701–707.
- Moody, C. J.; Roffey, J. R. A.; Stephens, M. A.; Stradford, I. J. Anti-Cancer Drugs 1997, 8, 489–499.
- Takasugi, M.; Monde, K.; Katsui, N.; Shirata, A. Chem. Lett. 1987, 1631–1632.
- Monde, K.; Sasaki, K.; Shirata, A.; Takasugi, M. *Phyto-chemistry* 1991, 30, 2915–2917.
- Suchý, M.; Kutschy, P.; Monde, K.; Goto, H.; Harada, N.; Takasugi, M.; Dzurilla, M.; Balentová, E. J. Org. Chem. 2001, 66, 3940–3947.

- Gross, D.; Porzel, A.; Schmidt, J. Z. Naturforsch., Sect. C 1994, 49, 281–285.
- 11. Monde, K.; Takasugi, M.; Shirata, A. *Phytochemistry* **1995**, *39*, 581–586.
- 12. For a review on 1-hydroxyindoles see: Somei, M. *Hetero-cycles* **1999**, *50*, 1157–1211.
- Monde, K.; Takasugi, M.; Ohnishi, T. J. Am. Chem. Soc. 1994, 116, 6650–6657.
- Takasugi, M.; Monde, K.; Katsui, N.; Shirata, A. Bull. Chem. Soc. Jpn. 1988, 61, 285–289.
- 15. Pedras, M. S. C.; Zaharia, I. L. *Phytochemistry* **2000**, *55*, 213–216.
- Kutschy, P.; Dzurilla, M.; Takasugi, M.; Török, M.; Achbergerová, I.; Homzová, R.; Rácová, M. *Tetrahedron* 1998, 54, 3549–3566.
- 17. Data for 14a: White solid, mp 72-74°C (CHCl₃/light petroleum); R_f (benzene/acetone, 19:1) 0.21; [Found: C, 54.4; H, 5.5; N, 8.2. C₁₆H₂₀N₂O₃S₂ requires C, 54.52; H, 5,72; N, 7.95]; v_{max} (CHCl₃) 3607, 2993, 2940, 2880, 1687, 1570, 1480, 1370, 1163 cm⁻¹; $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.59 (1H, d, J 8.1 Hz, H-7), 7.42 (1H, dd, J 7.6, 0.7 Hz, H-4), 7.28 (1H, ddd, J 7.9, 7.9, 1.4 Hz, H-6), 7.04 (1H, ddd, J 7.4, 7.4, 1.2 Hz, H-5), 6.95 (1H, d, J 6.3 Hz, OH-D₂O exchangeable), 5.63 (1H, d, J 6.4 Hz, CH), 4.83 (1H, d, J 15.6 Hz, H_b), 4.38 (1H, d, J 15.9 Hz, H_a), 2.54 (3H, s, SCH₃), 1.53 (9H, s, C(CH₃)₃); $\delta_{\rm C}$ (100 MHz, DMSO-d₆) 161.6 (C), 150.9 (C), 140.6 (C), 130.0 (C), 129.2 (CH), 123.6 (CH), 122.8 (CH), 114.2 (CH), 90.8 (CH), 81.0 (C), 70.6 (C), 65.9 (CH₂), 27.6 (CH₃), 14.3 (SCH₃); m/z (EIMS) 352 (15, M⁺), 296 (72), 252 (20), 234 (17), 161 (32), 145 (71), 117 (23), 87 (20), 57 (100). Data for 14b: Colourless plates, mp 126-128 °C (CH₂Cl₂/light petroleum); R_f (benzene/acetone, 19:1) 0.29; [Found: C, 54.3; H, 5.9; N, 8.1. C₁₆H₂₀N₂O₃S₂ requires C, 54.52; H, 5,72; N, 7.95]; v_{max} (CHCl₃) 3587, 3013, 2986, 2940, 1683, 1560, 1480, 1366, 1163 cm⁻¹; $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.61 (1H, d, J 7.8 Hz, H-7), 7.29-7.24 (2H, m, H-4, H-6), 7.05-7.00 (2H, m, H-5, OH-D₂O exchangeable), 5.49 (1H, d, J 6.0 Hz, CH), 4.36 (1H, d, J 15.4 Hz, H_b), 3.82 (1H, d, J 15.1 Hz, H_a), 2.49 (3H, s, SCH₃), 1.54 (9H, s, C(CH₃)₃); δ_C (100 MHz, DMSO-*d*₆) 164.1 (C), 151.0 (C), 139.2 (C), 132.1 (C), 128.8 (CH), 132.2 (CH), 122.6 (CH), 114.1 (CH), 87.6 (CH), 80.9 (C), 74.4 (CH₂), 73.7 (C), 27.6 (CH₃), 14.2 (SCH₃); EIMS identical with 14a.
- 18. Kawasaki, T.; Somei, M. *Heterocycles* **1990**, *31*, 1605–1608.
- 19. Somei, M.; Kobayashi, K.; Shimizu, K.; Kawasaki, T. *Heterocycles* **1992**, *33*, 77–80.
- 20. Mosmann, T. J. Immunol. Methods 1983, 65, 155-163.
- 21. Wattenberg, L. W. Cancer Res. 1992, 52, 2085-2091.